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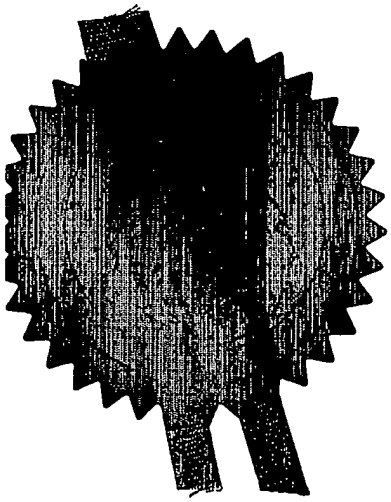

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Patents ADP number *(if you know it)* 8497925001

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4. Title of the invention — A DIAGNOSTIC SYSTEM FOR CARRYING OUT A NUCLEIC ACID SEQUENCE AMPLIFICATION AND DETECTION PROCESS

5. Name of your agent *(if you have one)* — BOULT WADE TENNANT

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A diagnostic system for carrying out a nucleic acid sequence amplification and detection process

The present invention is concerned with nucleic acid (NA) extraction and, in particular, an integrated lab-on-a-chip diagnostic system for carrying out a NA sequence amplification and detection process on a fluid sample containing cells.

To achieve both purification and preconcentration, analytical chemists have generally resorted to some kind of extraction procedure. These methods involve removal of the analytes of interest from the sample matrix, or alternatively, removing all other species from the sample matrix to leave behind the analytes of interest. Extraction processes can involve transfer of species from one liquid phase to another, or the capture of species from a liquid phase onto a solid surface. In the former case, preconcentration of a species is generally not achieved, unless solvent is actively removed from the phase containing that species. In the latter case, however, preconcentration can be achieved, if (a) the available binding area is large enough to bind more molecules than are present in the solution in contact with the surface at any one time, and (b) species can be efficiently removed from the solid phase using only a small amount of eluent. Since preconcentration is an important aspect of the nucleic acid sample pre-treatment procedure, solid-phase extraction has been adopted. A well-established nucleic acid extraction method involving binding of DNA to silica particles in the presence of a chaotropic agent (see Boom et al, J. Clin. Microbiol. 1990, 28, 495-503). The present invention involves

The isolation and purification of DNA and/or RNA from bacterial cells and virus particles is a key step in many areas of technology such as, for example, diagnostics, environmental monitoring, forensics and molecular biology research.

Microfabrication is an attractive construction method for producing devices for carrying out biological processes for which very small sample volumes are desirable, such as DNA sequence analysis and detection.

One such device, for carrying out a polymerase chain reaction (PCR) followed by a detection step is disclosed in US 5,674,742. Lamb wave pumps are used to transport DNA primers, polymerase reagents and nucleotide reagents from three separate storage chambers into a single reaction chamber as and when required to carry out a PCR process, with the temperature of the reaction chamber being cycled as required.

Another microfabricated device, for carrying out a chemical reaction step followed by an electrophoresis separation step, is disclosed in Analytical Chemistry 1994, 66, 4127-4132. Etched structures in a silicon substrate covered by a glass plate provide a reaction chamber and connections to buffer, analyte, reagent and analyte waste reservoirs, as well as an electrophoresis column connected to a waste reservoir.

Nucleic acid sequence-based amplification (NASBA) is a primer-dependent technology that can be used for the continuous amplification of nucleic acids in a single

internal standards are mixed with the wild-type nucleic acid after its isolation from the clinical sample. Quantitative NASBA is discussed in Nucleic Acid Research (1998) volume 26, pages 2150-2155. Post-NASBA product detection, however, can still be a labour-intensive procedure, normally involving enzymatic bead-based detection and electrochemiluminescent (ECL) detection or fluorescent correlation spectrophotometry. However, as these methodologies are heterogeneous or they require some handling of sample or robotic devices that are currently not cost-effective they are relatively little used for high-throughput applications. A homogeneous procedure in which product detection is concurrent with target amplification by the generation of a target-specific signal would facilitate large-scale screening and full automation. Recently, a novel nucleic acid detection technology, based on probes (molecular beacons) that fluoresce only upon hybridization with their target, has been introduced.

Fluidics is the science of liquid flow in, for example, tubes. For microfabricated devices, flow of a fluid through the one or more sets of micro or nano sized reaction chambers is typically achieved using a pump such as a syringe, rotary pump or precharged vacuum or pressure source external to the device. Alternatively, a micro pump or vacuum chamber, or lamb wave pumping elements may be provided as part of the device itself. Other combinations of flow control elements including pumps, valves and precharged vacuum and pressure chambers may be used to control the flow of fluids through the reaction chambers. Other mechanisms for transporting fluids within the system include electro-osmotic flow.

The present invention provides a system for carrying out a nucleic acid sequence amplification and detection process on a fluid sample containing cells and/or particles, the system comprising:

(a) an inlet for a fluid sample;

(b) a lysis unit for lysis of cells and/or particles contained in the fluid sample;

(c) a nucleic acid extraction unit for extraction of nucleic acids from the cells and/or particles contained in the fluid sample;

(d) a reservoir containing a lysis fluid;

(e) a reservoir containing an eluent for removing nucleic acids collected in the nucleic acid extraction unit;

wherein the sample inlet is in fluid communication with the lysis unit, an optional valve being present to control the flow of fluid therebetween;

wherein the lysis unit is in fluid communication with the nucleic acid extraction unit, an optional valve being present to control the flow of fluid therebetween;

wherein the reservoir containing the lysis fluid is in fluid communication with the lysis unit, an optional valve being present to control the flow of fluid therebetween; and

wherein the reservoir containing the eluent is in fluid communication with the nucleic acid extraction unit, an optional valve being present to control the flow of fluid therebetween.

The system can be used on millilitre sample volumes for routine diagnostics. The system relies on certain reagents being pre-loaded.

The system involves concentration of, for example, infected epithelial cells, lysis and extraction of mRNA, and real-time amplification and detection.

The system may be used for the screening of cervical carcinoma, for example.

At least some of the components of the system are preferably microfabricated. Preferably, the lysis unit, the nucleic acid extraction unit, the lysis fluid reservoir and the eluent reservoir are microfabricated and integrated, i.e. formed on a common substrate.

The reservoir containing the lysis fluid is preferably in fluid communication with the inlet, an optional valve being present to control the flow of fluid therebetween.

The reservoir containing the eluent is preferably in fluid communication with the inlet, an optional valve being present to control the flow of fluid therebetween.

The system will typically further comprise (g) a nucleic acid sequence amplification and detection unit, wherein the nucleic acid extraction unit is in fluid communication with the nucleic acid sequence amplification and detection unit, an optional valve being present to control the flow of fluid therebetween. Preferably, the nucleic acid sequence amplification and detection unit is microfabricated and preferably integrated with the other components.

More advantageously, the eluent, the first washing solvent (eg ethanol) and/or the second washing solvent (eg isopropanol) are contained in a common reservoir. This may be achieved by separating the eluent, the first washing solvent and/or the second washing solvent from one another in the common reservoir by the use of a fluid such as, for example, air. Other "separating" fluids (liquids or gases) can be used, however, as long as they are immiscible or at least substantially immiscible with the eluent, the first washing solvent and/or the second washing solvent.

In a preferred embodiment, the eluent, the ethanol and/or the isopropanol are contained in a conduit or channel which is in fluid communication with the inlet and the lysis unit. The eluent, the ethanol and/or the isopropanol being separated by fluid gaps such as air gaps, for example.

The system will typically further comprise (k) means for introducing a fluid sample and/or air into the inlet. Said mean preferably comprising a pump or a syringe. Alternatively, such means may comprises one or more variable volume chambers in communication with the inlet port, wherein altering the volume of the variable volume chamber(s) effects and/or restricts flow of a fluid sample into and/or out of the inlet. The variable volume chamber typically comprises a flexible membrane overlying a hollow recess in the underlying substrate.

The system may advantageously be driven by a single pumping system.

The lysis unit may have any suitable shape and configuration but will typically be in the form of a channel or chamber. The lysis unit is preferably for lysis of eukaryotic and prokaryotic cells and particles contained in the fluid sample.

If desired, the system may further comprise a filtration unit, which unit is in fluid communication with the lysis unit. The filtration unit may comprise, for example, a cross-flow filter or a hollow filter. Alternatively, the lysis unit may itself further comprise means to filter the fluid sample. Said mean may comprise, for example, a cross-flow filter or a hollow filter, which may be integrated with the lysis unit.

If desired, the system may further comprise a fragmentation unit, which unit is in fluid communication with the lysis unit. Alternatively, the lysis unit may itself further comprise means to fragment the fluid sample. Random fragmentation of DNA or RNA is often necessary as a sample pre-treatment step. Fragmentation may be achieved biochemically using restriction enzymes, or through application of a physical force to break the molecules (see, for example, P. N. Hengen, Trends in Biochem. Sci. , vol. 22, pp. 273- 274, 1997 and P. F. Davison, Proc. Nat. Acad. Sci. USA , vol. 45, pp. 1560- 1568, 1959). DNA fragmentation by shearing usually involves passing the sample through a short constriction. In a preferred embodiment, DNA and/or RNA breaks under mechanical force when pumped through a narrow orifice, due to rapid stretching of the molecule. A pressure-driven flow can lead

to a shear force, which leads to fragmentation of the nucleic acids.

The lysis unit may itself further comprise means to filter the fluid sample and means to fragment the fluid sample.

The system may further comprises means for heating the contents of the lysis unit and/or the nucleic acid extraction unit. Said mean may comprise, for example, one or more Peltier elements located in or adjacent the lysis unit and/or the nucleic acid extraction unit.

The nucleic acid extraction unit may have any suitable shape and configuration but will typically be in the form of a channel or chamber. The nucleic acid extraction unit is preferably for extraction of eukaryotic and prokaryotic cells and particles contained in the fluid sample.

The nucleic acid extraction unit may be at least partially filled with silica beads or particles. One or more sets of electrodes may be provided adjacent the silica beads or particles for collecting and/or pre-concentrating the eluted nucleic acids. The one or more sets of electrodes may comprise platinum electrodes, for example. Means may therefore be provided for applying a potential difference across the electrodes. The extraction cell is preferably formed from or comprises poly(dimethylsiloxane) (PDMS). The unit will typically comprise a substrate and an overlying cover, the extraction unit being defined by a recess in a surface of the substrate and the adjacent surface of the cover. The substrate is preferably formed

from silicon poly(dimethylsiloxane) (PDMS). The NA binds to silica surfaces in the presence of chaotropic agents.

The integration of electrodes (eg platinum electrodes) may advantageously be used to reversibly collect and pre-concentrate the eluted NA on-chip. Thus, the present invention enables combined nucleic acid extraction and enrichment to be achieved.

In a preferred embodiment, the nucleic acid extraction unit comprises a silica bead-packed poly(dimethylsiloxane) (PDMS) channel.

The system or at least a master version thereof will typically be formed from or comprise a semiconductor material, although dielectric (eg glass, fused silica, quartz, polymeric materials and ceramic materials) and/or metallic materials may also be used. Examples of semiconductor materials include one or more of: Group IV elements (i.e. silicon and germanium); Group III-V compounds (eg gallium arsenide, gallium phosphide, gallium antimonide, indium phosphide, indium arsenide, aluminium arsenide and aluminium antimonide); Group II-VI compounds (eg cadmium sulphide, cadmium selenide, zinc sulphide, zinc selenide); and Group IV-VI compounds (eg lead sulphide, lead selenide, lead telluride, tin telluride). Silicon and gallium arsenide are preferred semiconductor materials. The system may be fabricated using conventional processes associated traditionally with batch production of semiconductor microelectronic devices, and in recent years, the production of semiconductor micromechanical devices. Such microfabrication technologies include, for example,

epitaxial growth (eg vapour phase, liquid phase, molecular beam, metal organic chemical vapour deposition), lithography (eg photo-, electron beam-, x-ray, ion beam-), etching (eg chemical, gas phase, plasma), electrodeposition, sputtering, diffusion doping, ion implantation and micromachining. Non-crystalline materials such as glass and polymeric materials may also be used.

Examples of polymeric materials include PMMA (Polymethyl methylacrylate), COC (Cyclo olefin copolymer), polyethylene, polypropylene, PL (Polylactide), PBT (Polybutylene terephthalate) and PSU (Polysulfone), including blends of two or more thereof. The preferred polymer is PDMS or COC.

The device/system will typically be integrally formed. The device/system may be microfabricated on a common substrate material, for example a semiconductor material as herein described, although a dielectric substrate material such as, for example, glass or a ceramic material could be used. The common substrate material is, however, preferably a plastic or polymeric material and suitable examples are given above. The system may preferably be formed by replication of, for example, a silicon master.

The advantages of using plastics instead of silicon-glass for miniaturized structures are many, at least for biological applications. One of the greatest benefits is the reduction in cost for mass production using methods like microinjection moulding, hot embossing and casting. A factor of a 100 or more is not unlikely for complex structures. The possibility to replicate structures for

multilayered mould inserts gives a great flexibility of design freedom. Interconnection between the micro and macro world are in many cases easier because one got the option to combine standard parts normally used. Different approaches can be used for assembly techniques, like e.g. US-welding with support of microstructures, laser welding, gluing and lamination. Other features that are profitable is surface modification. For miniaturized structures addressed for biological analysis, it is important that the surface is biocompatible. By utilizing plasma treatment and plasma polymerization a flexibility and variation of assortment can be adapted into the coating. Chemical resistance against acids and bases are much better for plastics than for silicon substrates that are easily etched away. Most detection methods within the biotechnological field involves optical measurements. The transparency of plastic is therefore a major feature compared to silicon that are not transparent. Polymer microfluidic technology is now an established yet growing field within the Lab-on-a-chip market.

The microfabricated system as herein described is also intended to encompass nanofabricated devices.

For a silicon or semiconductor master, it is possible to define by, for example, etching or micromachining, one or more of variable volume chambers, microfluidic channels, reaction chambers and fluid interconnects in the silicon substrate with accurate microscale dimensions. A plastic replica may then be made of the silicon master. In this manner, a plastic substrate with an etched or machined

microstructure may be bonded by any suitable means (for example using an adhesive or by heating) to a cover.

The optional valves used in the system may take any convenient form. For example, the valves may simply regulate flow along a conduit or channel connecting two units. A piston-like member may be provided which can be raised or lowered in a hole in a conduit or channel by the action of a pin device.

Use of the system involves the following possible steps, by way of example.

Alternative 1

(i) Sample collection and lysis
(ii) Extraction of mRNA (manual or automatic procedure)
(iii) Real-time amplification and detection (preferably multiplex)

Alternative 2

(iv) A fragmentation unit may include both sample lysis and sample preparation
(v) Real-time amplification (NASBA) and detection (preferably multiplex).

The present invention also provides a method for the manufacture of an integrated lab-on-a-chip diagnostic system as herein described, which method comprises:

A. providing a substrate having an inlet recess, a lysis unit recess, a nucleic acid extraction unit recess, a lysis

The eluent, and/or the ethanol and/or the isopropanol are preferably separated from one another by a fluid, preferably air, although any immisible fluid (liquid or gas) may be used.

In a preferred embodiment the method comprises:

introducing eluent into the eluent reservoir after bonding the cover to the substrate;

introducing a first volume of air into the eluent reservoir;

introducing ethanol into the eluent reservoir, whereby the ethanol is separated from the eluent by said first volume of air;

introducing a second volume of air into the eluent reservoir;

introducing isopropanol into the eluent reservoir, whereby the isopropanol is separated from the ethanol by said second volume of air.

The substrate may be formed from silicon, for example, and the overlying cover from glass, for example. In this case, the glass cover is preferably anodically bonded to the silicon substrate, optionally through an intermediate silicon oxide layer formed on the surface of the substrate. The recesses in the silicon may be formed using reactive-ion etching. Other materials such as polymeric materials may also be used for the substrate and/or cover. Such materials may be fabriacted using, for example, a silicon replica. Alternatively, the device may be fabricated by structuring of mould inserts by milling and electro-discharge machining (EDM), followed by injection moulding of the chip parts, followed by mechanical post-processing of the polymer parts,

for example drilling, milling, debarring. This may subsequently be followed by insertion of the filter, solvent bonding, and mounting of fluidic connections.

Examples of polymeric materials include PMMA (Polymethyl methylacrylate), COC (Cyclo olefin copolymer), polyethylene, polypropylene, PL (Polylactide), PBT (Polybutylene terephthalate) and PSU (Polysulfone), including blends of two or more thereof. COC is preferred.

Preferably, and in particular if optical observations of the contents of the cell are required, the overlying cover is made of an optically transparent substance or material, such as glass, Pyrex or COC.

Combinations of a microfabricated component with one or more other elements such as a glass plate or a complementary microfabricated element are frequently used and intended to fall within the scope of the term microfabricated used herein.

Part or all of the substrate base may be provided with a coating of thickness typically up to 1 μm, preferably less than 0.5 μm. The coating is preferably formed from one or more of the group comprising polyethylene glycol (PEG), Bovine Serum Albumin (BSA), tweens and dextrans. Preferred dextrans are those having a molecular weight of 9,000 to 200,000, especially preferably having a molecular weight of 20,000 to 100,000, particularly 25,000 to 75,000, for example 35,000 to 65,000). Tweens (or polyoxyethylene sorbitans) may be any available from the Sigma Aldrich Company. PEGs are preferred as the coating means, either

singly or in combination. By PEG is embraced pure polyethylene glycol, i.e. a formula $HO-(CH_2CH_2O)_n-H$ wherein n is an integer whereby to afford a PEG having molecular weight of from typically 200 - 10,000, especially PEG 1,000 to 5,000; or chemically modified PEG wherein one or more ethylene glycol oligomers are connected by way of homobifunctional groups such as, for example, phosphate moieties or aromatic spacers. Particularly preferred are polyethylene glycols known as FK108 (a polyethylene glycol chain connected to another through a phosphate); and the PEG sold by the Sigma Aldrich Company as product P2263. The above coatings applied to the surfaces of the cell/chamber, inlets, outlets, and/or channels can improve fluid flow through the system. In particular, it has been found that the sample is less likely to adhere or stick to such surfaces. PEG coatings are preferred.

For a silicon or semiconductor master, it is possible to define by, for example, etching or micromachining, one or more of variable volume chambers, microfluidic channels, reaction chambers and fluid interconnects in the silicon substrate with accurate microscale dimensions (deep reactive-ion etching (DRIE) is a preferred technique). A plastic replica may then be made of the silicon master. In this manner, a plastic substrate with an etched or machined microstructure may be bonded by any suitable means (for example using an adhesive or by heating) to a cover thereby forming the enclosed fragmentation cell(s), inlet(s), outlet(s) and connecting channel(s).

The device comprises a substrate with the desired microstructure formed in its upper surface. The substrate

break down the bacterial cell wall or virus protein coating and isolate nucleic acids. This is important when the starting material consists of a large volume, for example an aqueous solution containing relatively few bacterial cells or virus particles. This type of starting material is commonly encountered in environmental testing applications such as the routine monitoring of bacterial contamination in drinking water.

The system is preferably designed to cater for a sample volume of 10-100 ml.

The present invention also provides an apparatus for the analysis of biological and/or environmental samples, the apparatus comprising a system as herein described. The apparatus may be a disposable apparatus.

The present invention also provides an assay kit for the analysis of biological and/or environmental samples, the kit comprising a system as herein described and means for contacting the sample with the system. The assay kit may be a disposable kit.

The present invention will now be described, by way of example, with reference to the accompanying drawings, of which:

Figure 1 is a schematic illustration of a sandwich layout used for integration of a flat membrane into a disposable polymer chip device for use in the present invention.

Figure 2 is a schematic illustration of a valve design for use with the system according to the present invention.

Figure 3 is a schematic illustration of a valve design for use with the system according to the present invention.

Figure 4 is a schematic illustration a possible layout of a bead chamber according to the present invention.

Figure 5 is a schematic illustration of a system design according to the present invention showing filling with lysis buffer and extraction fluids.

Figure 6 is a schematic illustration of a chip layout according to a preferred embodiment of the present invention.

Figure 7 is a schematic illustration of a system design according to another preferred embodiment of the present invention.

Figure 8 relates to the Examples.

Figure 9 relates to the Examples.

A plastic chip design according to the present invention preferably incorporates supply channels, reaction chambers and microfluidic actuation systems and is preferably processed by injection moulding of cycloolefin copolymer (COC). The mould insert for, for example, a 12-channel chip may be manufactured using high precision milling. The detection volume is typically approximately 80

nL (400 x 2000 x 100 μm). The plastic chip is preferably first oxygen plasma activated before being coated with a 5% polyethylene glycol (PEG) solution (Sigma Chemical Co, St. Louis, MO). After coating, the chip may be sealed with an approximately 75 μm COC membrane via solvent welding using, for example, bicyclohexcyl. A thin gold layer (approx 25 nm) is preferably deposited on the backside of the chip to prevent background fluorescence from the thermal pad on top of the Peltier element.

If required, Peltier elements may be integrated into the sample holder providing thermal control for the plastic chips. Aluminium blocks may be put on top of the Peltier elements to secure an even distribution of heat for the chips. A thermal pad is preferably mounted on the aluminium blocks to establish thermal contact between the chips and the heating source. A thermocouple will typically be placed on the sample holder measuring the air temperature and having a feedback circuit to the Peltier elements. The temperature regulation can be controlled externally on a laptop.

As previously described, NASBA is an isothermal (approximately 41°C) amplification method specifically designed for amplifying any single-stranded RNA sequence. The NASBA reaction can be applied to a wide range of applications such as detection of the presence of specific viral RNAs, RNAs of other infectious or pathogenic agents or certain cellular RNAs. Simultaneous activity of the three enzymes, AMV Reverse Transcriptase, RNase H and T7 RNA polymerase makes the core technology in the amplification reaction. Two oligonucleotide primers determine the

specificity of the reaction and fluorescent molecular beacon probes that are specific for the target RNA. In approximately 90 minutes the nucleic acid sequence of interest can be amplified to > 10^9 copies. The optical detection unit is preferably designed to excite the fluorophores in the reaction chambers at approximately 494 nm and detect the emitted fluorescent light at approximately 525 nm. The excitation light may be filtered using a bandwidth filter (465 nm - 500 nm) before the light is collimated through a lens. The same Fresnel lens may be used for focusing the illumination and collection of the fluorescence light. Another lens may be used to focus the fluorescent light onto the detector surface (eg a photomultiplier-tube). The data collection and preparation of the detected signal may be processed on a laptop using MATLAB 6.0.088 Release 12 (The MathWorks Inc., Natick, MA).

Efficient sample pre-treatment is an important factor in the context of micro-technological analysis systems. In particular, concentration devices are needed in order to enable detection of low numbers of specific particles, as e.g. cells bacteria or viruses, present in biological samples. A variety of concentration methods are known in the art including, for example, filtration techniques such as dead-end filtration and cross-flow filtration using different kinds of filtration media (micro-structured channels, porous hollow fibres or membranes), gravity settlers, centrifuges, acoustic cell filters, optical traps, dielectrophoresis (DEP), electrophoresis, flow cytometry and adsorption based methods.

First, the cell suspension is applied to the filtration unit by means of a syringe pump. Besides particulate suspension the syringe is loaded with about 200 µL to 300 µL of air, which is used for actuation of the on-chip liquids (Depending on the application it will be appreciated that other immiscible liquids may be used).

Second, air is pumped into the lysis buffer reservoir and the displaced buffer is applied to the cells being kept on the filter. The cell lysate is pushed through the filter and is directed to the beads chamber. Due to the additional filtering step the probability of clogging in the beads chamber is reduced.

Third, the actuation pump (syringe) is connected to the extraction liquid reservoir while the connections to the filter chamber and the lysis buffer reservoir are closed. The extraction liquids are stored in a single reservoir separated by air plugs. When pressure is applied to one side of the reservoir, the liquids are displaced in parallel and are sequentially guided through the beads chamber.

The operation protocol including the valve operations is summarized below. Valves not listed are in a closed state, whereas the listed valves are opened for the corresponding operation.

Filtration

Valves 5, 7: Cell suspension in, filtrate -> Left Outlet

Lysis

Valves 2, 3, 7:	Air in, displaced fluid -> Left Outlet
Valves 2, 3, 6:	Air in, lysate -> bead package, Right Outlet

Purification

Valves 1, 4, 6:	Air in, isopropanol -> bead package
	Air in, ethanol -> bead package
	Air in, elution buffer -> bead package

Turning now to Figure 7, which shows another preferred embodiment of the present invention. The foregoing description is also applicable to this embodiment. The system 1 comprises an inlet 5 for a fluid sample, a lysis/filtration unit 10, a nucleic acid extraction unit 15, a channel 20 containing lysis fluid, a channel 25 containing eluent, ethanol and isopropanol, a nucleic acid sequence amplification and detection unit 30, and a waste unit 35.

A channel 11 connects the sample inlet 5 to the lysis/filtration unit 10. A valve 12 is provided to control the flow of fluid therebetween.

A channel 16 connects the lysis/filtration unit 10 to the nucleic acid extraction unit 15. A valve 17 is provided to control the flow of fluid therebetween.

The channel 20 containing the lysis fluid is connected to the lysis/filtration unit 10 and the sample inlet 5. Valve 22s and 23 are provided to control the flow of fluid.

The channel 25 containing the eluent, ethanol and isopropanol is connected to the nucleic acid extraction unit 15 and the sample inlet 5. Valves 27 and 28 are provided to control the flow of fluid.

A channel 31 connects the nucleic acid extraction unit 15 to the nucleic acid sequence amplification and detection unit 30. A valve 32 is provided to control the flow of fluid therebetween.

A channel 36 connects the lysis/filtration unit 10 to the waste unit 35. A valve 37 is provided to control the flow of fluid therebetween.

The channel 25 contains the eluent and washing solvents such as ethanol and isopropanol. The eluent and washing solvents are preloaded into the channel using an air gap to separate the liquids from one another.

An example of a suitable lysis buffer fluid is 100 mM Tris/HCl, 8 M GuSCN (pH 6.4).

An example of a suitable elution solution is 10 mM Tris/HCl, 1 mM EDTA Na_2 (pH 8) + 1 mM YOYO-1.

Nucleic acid quantification may be achieved using a fluorescence microscope and a pixel-intensity analysis program (Lispix).

The nucleic acid extraction unit contains silica beads, for example 0.3 mg of 15-30 µm size silica beads. Platinum elctrodes are also provided (not shown) just below the

Purification/Extraction

All valves are closed except for valves 27, 28 and 32. In a first step, air contained in the syringe is injected into the sampele inlet 5. This causes the fluids (isopropanol, air gap, ethanol, air gap, elution buffer) contained in channel 25 to move as a column of fluid towards the nucleic acid extraction unit 15. This process is halted once all of the isopropanol (i.e. the first portion of the column of fluid) has been passed into the nucleic acid extraction unit 15. After a short period of time (together with optional heating of the contents of unit 15), the process is continued and the air gap between the isopropanol and the ethanol displaces the isopropanol. The isopropanol evaporates and/or goes to waste. The ethanol then flows under pressure into the nucleic acid extraction unit 15. The process is once again halted once all the ethonal has passed into the unit 15. After a short period of time (together with optional heating of the contents of unit 15), the process is continued and the air gap between the ethanol and the elution buffer displaces the ethanol. The ethanol evaporates and/or goes to waste. The elution buffer then flows under pressure into the nucleic acid extraction unit 15 and elutes the nucleic acids released from the surface of the silica beads. The eluted nucleic acids then pass to the nucleic acid sequence amplification and detection unit 30.

The present invention provides an apparatus and method for nucleic acid (NA) extraction and analysis. Extraction from biological samples, such as human cell lysates, has been successful, with collection of the NA in the first 15 mL of eluate.

Mantyjarvi, R., and Syrjanen, K. *J Virol Methods*, 1988, 19, 225 - 238). A ten-fold serial dilutions of the SiHa cell-line extract were tested. In addition, artificial HPV type 16 sequences, from the HPV Proofer kit (NorChip AS, Norway) was used as target. A dilution series were tested to define the detection limit of the system.

NASBA

The reagents in the PreTect® HPV-Proofer kit were mixed according to the manufacturers specifications (NorChip AS, Norway). All primers and probes were available in the kit. Additionally, BSA was added to the mixture to a final concentration of 0.05 % as a dynamic coating. Reagent solution (26 µL) from the kit and 13 µL of sample material (SiHa cell-line samples and HPV type 16 sequence samples from the kit) were mixed and heated to 65°C for 2 minutes. The mixture was subsequently cooled to 41°C for 2 minutes after which the enzymes (13 µL) were added. One actuation chamber on each reaction channel was cut open before adding the mixture into the polymer microchip. Each reaction channel in the chip was filled with the mixture due to capillary forces. The remaining mixture was drawn into the waste chamber at the end at the supply channel. The chip holder was then moved under the optics, where one after the other channel was measured. Measurements were taken every 30 seconds. Only a 2 x 2 mm^2 area were illuminated by the LED, this area corresponded to a detection area of 80 nL. The ten-fold serial dilutions of both HPV 16 sequences and SiHa cell-lines were also tested with conventional equipment for comparison with microchip detection. All experiments were run for 2.5 hours.

Calculation

All the results were calculated using PreTect Data Analyzer (PDA) (NorChip AS). The microchip was designed with 12 reaction chambers, but the two reaction channels on each side were removed in the calculations due to systematic error of the measurements. The calculations were based on polynomial regression algorithms. The ratio was defined as the difference in fluorescence level at the end of the reaction and the fluorescence level at the start of the reaction. All samples with a ratio of 1.7 or greater were defined to be positive. Time-to-positivity or the starting point were set to be where the curve started to increase exponentially. The average slop were calculated using the values of 10% increase in fluorescent level and the value of an 80% increase in fluorescent level from the starting point. The detection limit for the polymer microchips was set to be the last concentration tested where all the 10 reaction channels were positive.

Results

Identification of HPV 16 virus utilizing real-time NASBA was successfully performed in polymer microchips with a detection volume of 80 nL. Figures 8 and 9 illustrate the result from one experiment performed on SiHa cell-lines and HPV 16 oligo sequences, respectively. The Figures show graphs that clearly are positive and have the same curvature as samples performed using regular 20 µL volumes and conventional readers (not shown). Table 1 shows the results of a dilution series of artificial HPV16 sequences and SiHa cell-lines obtained using the polymer microchips. To characterize the amplification reactions, several different parameters were evaluated: the fluorescence ratio, time-to-

positivity, the average slop of the linear part of the curve, the number of positive amplifications and the number of polymer microchips tested. The values in the table show the average value and the standard deviation of the positive samples that were tested. For both HPV 16 sequences and SiHa cell lines tested on the microchips, the ratio was more or less constant. In comparison with conventional testing (Table 2) of the same sample material, showed that the ratio were decreasing for lower concentrations. The other parameters on the other hand correspond very well for both the microchips and the conventional methods. Time-to-positivity increased with lower concentrations. While the average slop values decreased with lower concentrations. Ten-fold serial dilutions from 100 aM to 100 nM were tested for artificial HPV 16 sequences, while SiHa cell-line were tested for 0.02 cells/µl to 2000 cells/µl. The custom-made optical detection system had a detection limit of 1 pM and 20 cells/µl for artificial HPV 16 sequences and SiHa cell-line material, respectively. These were the same detection limits obtained for the conventional Biotek readers. It was possible to detect lower concentrations on both systems but the results were not consistent. The results also illustrate that when the sample concentration of input target were decreasing, the standard deviation increased. A comparison of the NASBA results for both HPV 16 oligo sequences and SiHa cell lines showed that all parameters had the same trend for microsystems as well as for conventional methods except for the ratio between the levels of the fluorescence at the start and at the end of the amplification reaction. Background noise is more distinctive at small reaction chambers than for macroscopic fluorescence methods. Parts of the background fluorescence were removed from the assay

by applying a thin gold layer on the backside of the polymer microchips. The COC itself is autofluorescent always giving some background fluorescence. Another contribution to noise detection is light scattering due to less perfect polymer surfaces. Time-to-positivity decreased for lower concentration as expected because the substrates used longer times to find and interact with the substrates. For the highest concentrations especially for the artificial HPV 16 in the experiments the time-to-positivity increase. Very high sample concentrations may also inhibit the reaction and therefore use longer time than an ideal reaction mixture. In the same manners the average slope decreases. When smaller amounts of target are in the reaction mixture to begin with, less amplicons will be produced and the slope will become lower than for higher concentrations. The detection limit of the NASBA reaction depends on the target of interest, the design of the primers and probe. In these experiments we were able to detect concentrations down to 1 pM and 20 cells/µl in both detection systems. Accordingly, this Example shows that it is possible to detect artificial HPV 16 sequences down to 1 pM concentration in polymer microchips utilizing real-time NASBA. For cell-line samples the detection limit were 20 cells/µl. These detection limits are the same that were obtained for experiments performed in the conventional Biotek reader.

Table 1: NASBA performed on microchips detecting HPV 16 oligo sequences and SiHa cell-line dilution series. The results are the average and standard deviation of all values obtained in the experiments.

Concentration	Ratio	Start point	Average slope	Positive amplifications / Number of reactions	Number of chips tested
HPV 16 oligo sequence [µM]					
0.1	2.90 ± 0.33	12.31 ± 5.36	45.09 ± 9.89	50 / 50	5
0.01	3.06 ± 0.37	14.73 ± 4.03	43.48 ± 9.48	40 / 40	4
0.001	2.65 ± 0.42	9.00 ± 2.05	45.99 ± 17.66	30 / 30	3
0.0001	2.75 ± 0.32	22.19 ± 4.45	35.08 ± 17.94	30 / 30	3
0.00001	2.56 ± 0.38	22.55 ± 7.36	29.87 ± 13.74	30 / 30	3
0.000001	2.54 ± 0.46	25.30 ± 3.60	19.62 ± 9.21	30 / 30	3
0.0000001	2.10 ± 0.32	37.09 ± 12.74	17.27 ± 11.78	33 / 70	7
0.00000001	1.85 ± 0.28	43.75 ± 7.13	9.94 ± 3.55	6 / 60	6
0.000000001	2.27 ± 0.86	81.00 ± 38.18	15.02 ± 6.26	2 / 60	6
0.0000000001	3.93	4.50	21.83	1 / 60	6
SiHa cell line [cells/µl]					
2000	2.86 ± 0.30	16.91 ± 2.67	42.57 ± 6.24	40 / 40	4
200	2.80 ± 0.43	18.89 ± 3.39	40.56 ± 14.50	40 / 40	4
20	2.88 ± 0.27	30.65 ± 9.28	37.49 ± 11.09	39 / 40	4
2	2.75 ±0.50	38.02 ± 26.12	35.09 ± 15.47	60 / 70	7
0.2	2.73 ± 0.54	70.13 ± 39.12	39.29 ±14.97	4 / 50	5
0.02	0	0	0	0 / 30	3

Table 2: Conventional NASBA testing performed on HPV 16 oligo sequences and SiHa cell-lines. The results are the average and standard deviation of all values obtained in the experiments.

Concentration	Ratio	Start point	Average slope	Positive amplifications / Total reactions
HPV 16 oligo sequence [µM]				
0.1	6.51 ± 0.18	14.00 ± 0.77	111.21 ± 19.29	6 / 6
0.01	6.74 ± 0.27	11.75 ± 1.47	96.26 ± 28.28	6 / 6
0.001	6.47 ± 0.28	15.25 ± 1.75	113.05 ± 33.62	6 / 6
0.0001	5.18 ± 1.07	23.83 ± 4.65	94.42 ± 58.85	6 / 6
0.00001	4.80 ± 1.17	25.13 ± 3.68	84.10 ± 38.27	12 / 12
0.000001	3.84 ± 0.81	26.25 ± 5.52	42.68 ± 11.40	12 / 12
0.0000001	1.79 ± 0.09	33.75 ± 7.42	15.71 ± 1.53	2 / 12
0.00000001	-	-	-	0 / 12
0.000000001	-	-	-	0 / 12
0.0000000001	-	-	-	0 / 12
SiHa cell line [cells/µl]				
2000	4.85 ± 0.58	29.25 ± 1.25	80.09 ± 6.80	6 / 6
200	3.84 ±1.22	29.25 ± 4.00	52.47 ± 24.82	6 / 6
20	3.66 ± 1.15	33.30 ± 7.82	44.04 ± 16.82	5 / 6
2	2.96 ± 0.42	39.75 ± 1.06	27.95 ± 7.15	2 / 6
0.2	-	-	-	0 / 6
0.02	-	-	-	0 / 6

CLAIMS:

1. An integrated lab-on-a-chip diagnostic system for carrying out a sample preparation, nucleic acid sequence amplification and detection process on a fluid sample containing cells and/or particles, the system comprising:

(a) an inlet for a fluid sample;

(b) a lysis unit for lysis of cells and/or particles contained in the fluid sample;

(c) a nucleic acid extraction unit for extraction of nucleic acids from the cells and/or particles contained in the fluid sample;

(d) a reservoir containing a lysis fluid;

(e) a reservoir containing an eluent for removing nucleic acids collected in the nucleic acid extraction unit;

wherein the sample inlet is in fluid communication with the lysis unit, a valve being present to control the flow of fluid therebetween;

wherein the lysis unit is in fluid communication with the nucleic acid extraction unit, a valve being present to control the flow of fluid therebetween;

wherein the reservoir containing the lysis fluid is in fluid communication with the lysis unit, a valve being present to control the flow of fluid therebetween; and

wherein the reservoir containing the eluent is in fluid communication with the nucleic acid extraction unit, a valve being present to control the flow of fluid therebetween.

2. A system as claimed in claim 1, wherein the reservoir containing the lysis fluid is in fluid communication with the inlet, a valve being present to control the flow of fluid therebetween.

3. A system as claimed in claim 1 or claim 2, wherein the reservoir containing the eluent is in fluid communication with the inlet, a valve being present to control the flow of fluid therebetween.

4. A system as claimed in any one of claims 1 to 3, further comprising (g) a nucleic acid sequence amplification and detection unit, wherein the nucleic acid extraction unit is in fluid communication with the sample preparation and the nucleic acid sequence amplification and detection unit, a valve being present to control the flow of fluid therebetween.

5. A system as claimed in any one of claims 1 to 4, further comprising (h) a waste unit, wherein the waste unit is in fluid communication with the lysis unit, a valve being present to control the flow of fluid therebetween.

6. A system as claimed in any one of claims 1 to 5, further comprising (i) a reservoir containing a washing solvent (preferably ethanol), which reservoir is in fluid communication with the nucleic acid extraction unit, a valve being present to control the flow of fluid therebetween.

7. A system as claimed in any one of claims 1 to 6, further comprising (j) a reservoir containing a further washing solvent (preferably isopropanol), which reservoir is in fluid communication with the nucleic acid extraction unit, a valve being present to control the flow of fluid therebetween.

8. A system as claimed in claim 6 or claim 7, wherein the reservoir containing the eluent is in fluid communication with the reservoir containing the first washing solvent (eg ethanol) and/or the reservoir containing the second washing solvent (eg isopropanol).

9. A system as claimed in claim 8, wherein the eluent, the first washing solvent (eg ethanol) and/or the second washing solvent (eg isopropanol) are contained in a common reservoir.

10. A system as claimed in claim 9, wherein the eluent, the first washing solvent (eg ethanol) and/or the second washing solvent (eg isopropanol) are separated from one another in the common reservoir by a fluid, preferably air.

11. A system as claimed in claim 9 or claim 10, wherein the common reservoir comprises a conduit in fluid communication with the inlet and the lysis unit.

12. A system as claimed in any one of claims 1 to 11, further comprising (k) means for introducing a fluid sample and/or air into the inlet, said mean preferably comprising a pump or a syringe.

13. A system as claimed in any one of claims 1 to 11, further comprising a filtration unit, which unit is in fluid communication with the lysis unit.

14. A system as claimed in claim 13, wherein the filtration unit comprises one or more of a dead-end filter, a cross-flow filter (eg micro-structured channels, porous hollow

fibres or membranes), a gravity settler, a centrifuge, an acoustic cell filter, an optical trap, dielectrophoresis (DEP), electrophoresis, flow cytometry and adsorption based methods.

15. A system as claimed in any one of claims 1 to 11, wherein the lysis unit further comprises means to filter the fluid sample.

16. A system as claimed in claim 15, wherein said means comprises one or more of a dead-end filter, a cross-flow filter (eg micro-structured channels, porous hollow fibres or membranes), a gravity settler, a centrifuge, an acoustic cell filter, an optical trap, dielectrophoresis (DEP), electrophoresis, flow cytometry and adsorption based methods.

17. A system as claimed in any one of the preceding claims, wherein the system further comprises means for heating the contents of the lysis unit and/or the nucleic acid extraction unit.

18. A system as claimed in claim 17, wherein said mean comprises one or more Peltier elements located in or adjacent the lysis unit and/or the nucleic acid extraction unit.

19. A system as claimed in any one of the preceding claims, wherein the nucleic acid extraction unit is at least partially filled with silica beads or particles.

20. A system as claimed in claim 19, wherein further comprising one or more sets of electrodes adjacent the silica beads or particles for collecting and/or preconcentrating the eluted nucleic acids.

21. A system as claimed in claim 20, wherein said one or more sets of electrodes comprises platinum electrodes.

22. A system as claimed in any one of the preceding claims for extracting nucleic acids present in a biological fluid, a dairy product, an environmental fluid or drinking water.

23. An apparatus for the analysis of biological and/or environmental samples, the apparatus comprising a system as defined in any one of the preceding claims.

24. An assay kit for the analysis of biological and/or environmental samples, the kit comprising a system as defined in an one of the claims 1 to 22 and means for contacting the sample with the system.

25. An apparatus as claimed in claim 23 or an assay kit as claimed in claim 24 which is disposable.

26. A method for the manufacture of an integrated lab-on-a-chip diagnostic system as defined in any one of the preceding claims, which method comprises:

A. providing a substrate having an inlet recess, a lysis unit recess, a nucleic acid extraction unit recess, a lysis fluid reservoir recess and an eluent reservoir recess in a surface thereof;

B. providing a cover; and

32. A method as claimed in claim 26 or claim 27, further comprising:

introducing eluent into the eluent reservoir after bonding the cover to the substrate;

introducing a first volume of an immiscible fluid (eg air) into the eluent reservoir;

introducing a first washing solvent (eg ethanol) into the eluent reservoir, whereby the first washing solvent is separated from the eluent by said first volume of immiscible fluid;

introducing a second volume of immiscible fluid (eg air) into the eluent reservoir; and

introducing a second washing solvent (eg isopropanol) into the eluent reservoir, whereby the second washing solvent is separated from the first washing solvent by said second volume of immiscible fluid.

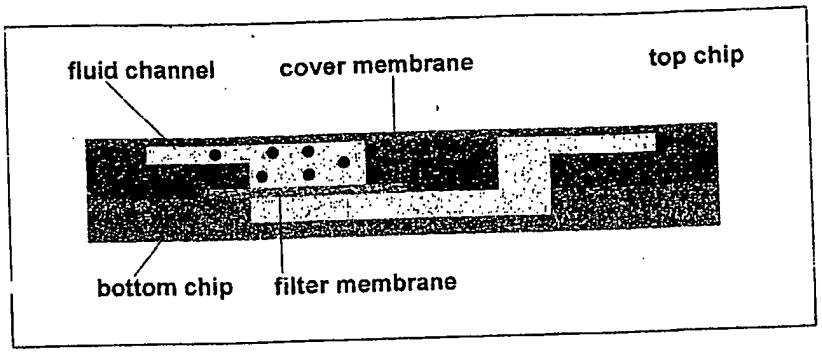

Figure 1: Schematic of the sandwich layout used for integration of a flat membrane into a disposable polymer chip

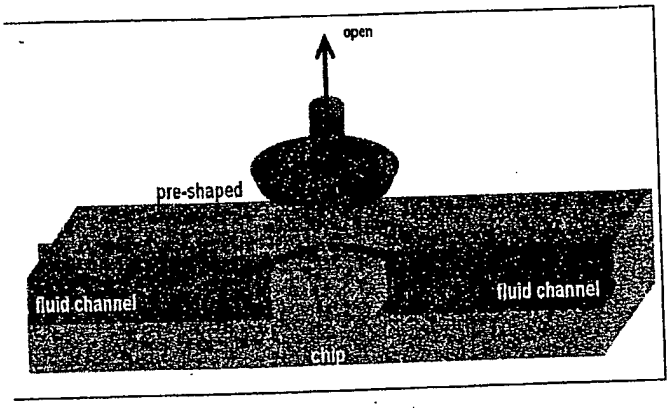

Figure 2: Functional principle of the disposable valve used for on-chip fluidic control.

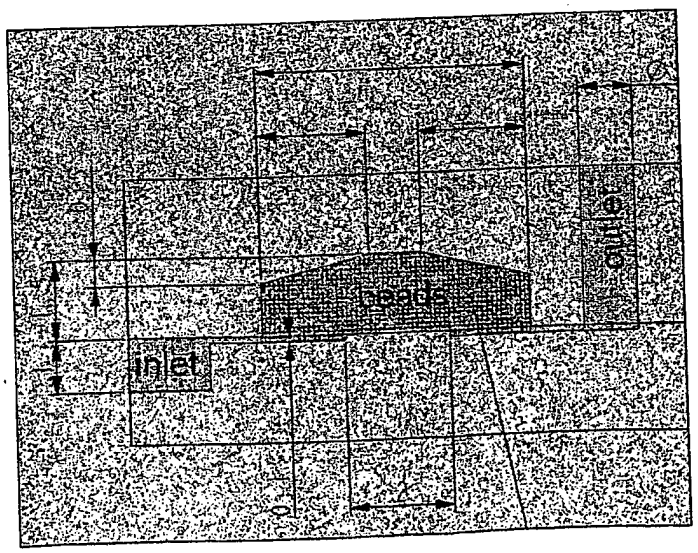

Figure 4: Beads chamber layout. Channel depth: 1 mm; Volume of beads chamber (dark blue): 6.5 µL

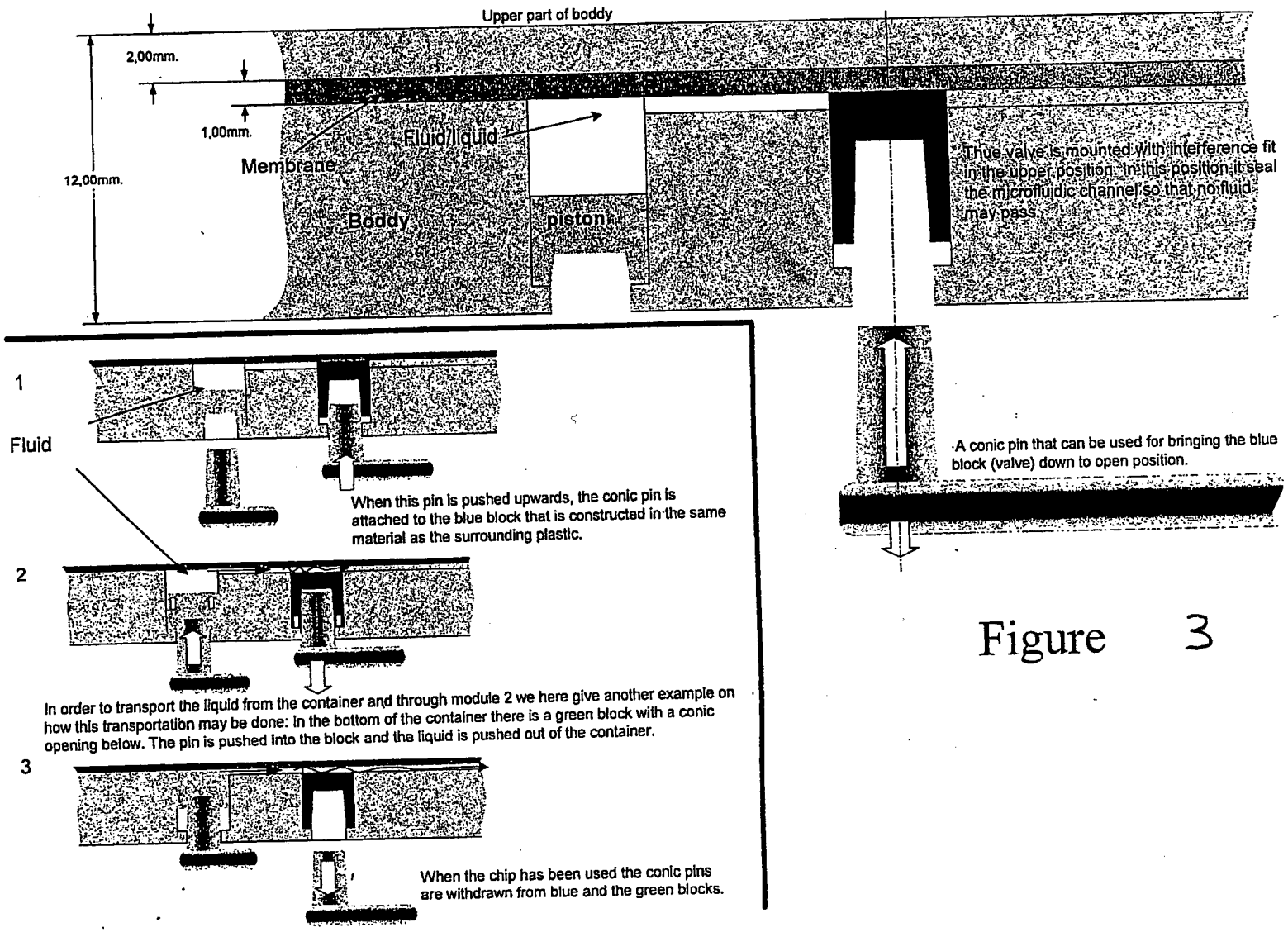
Upper part of boddy
2,00mm.
1,00mm.
12,00mm.
Membrane
Fluid/liquid
Boddy
piston
The valve is mounted with interference fit in the upper position. In this position it seal the microfluidic channel so that no fluid may pass.
A conic pin that can be used for bringing the blue block (valve) down to open position.
1
Fluid
When this pin is pushed upwards, the conic pin is attached to the blue block that is constructed in the same material as the surrounding plastic.
2
In order to transport the liquid from the container and through module 2 we here give another example on how this transportation may be done: In the bottom of the container there is a green block with a conic opening below. The pin is pushed into the block and the liquid is pushed out of the container.
3
When the chip has been used the conic pins are withdrawn from blue and the green blocks.

Figure 3

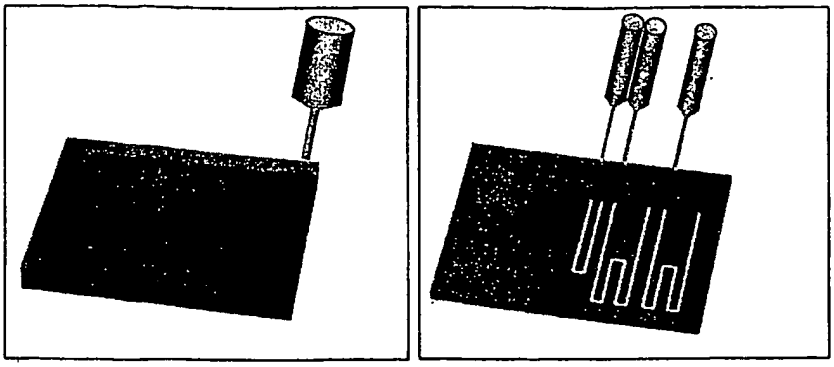

Figure 5: Left: top chip comprising the meandering reservoir for storage of lysis buffer.
Right: lower chip part comprising three reservoirs for storage of the extraction liquids. The positions of the filling channels are indicated by the schematic drawings of syringes.

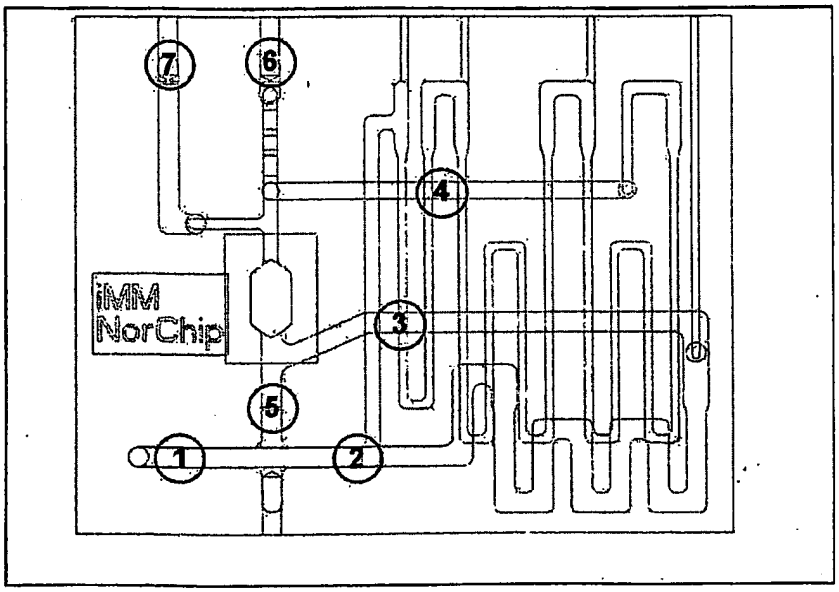

Figure 6: Chip layout. The valves (all located at the upper side of the top chip) are labelled with numbers. The meandering channels denote the liquid reservoirs as shown in Fig. 3-4. The central sexangle denotes the filtration chamber which is surrounded by a flat cut out on the lower side of the top chip used for a tight insertion of the filter membrane. The beads chamber, a cross section of which is shown in Fig. 3-3 is marked with the horizontal arrow. Circles denote channel connections between both chip parts. In- and outlet ports are denoted with vertical arrows.

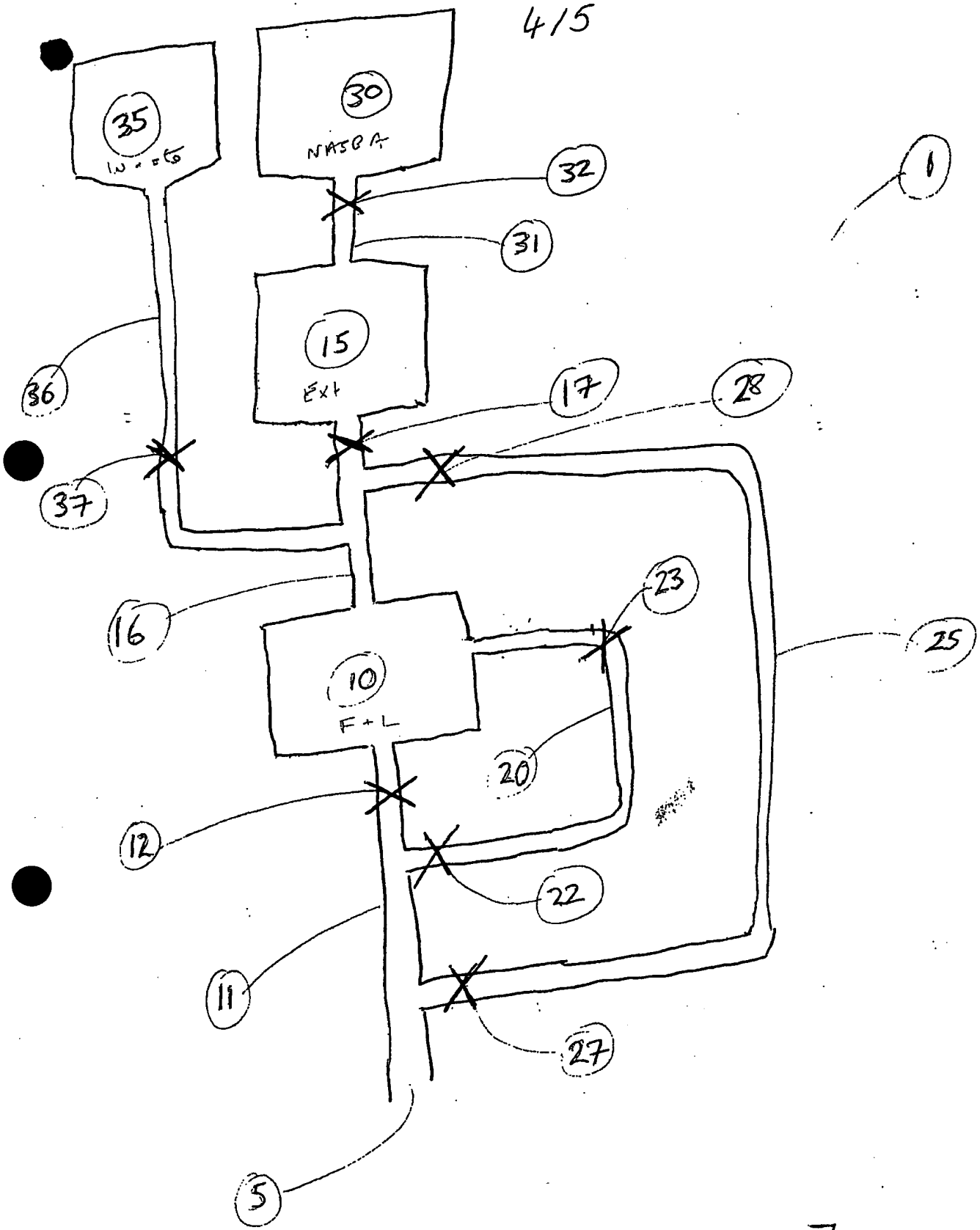
35
IN + EG
30
NASBA
32
1
31
15
EXT
36
17
28
37
16
23
25
10
F + L
20
12
22
11
27
5

Figure 7

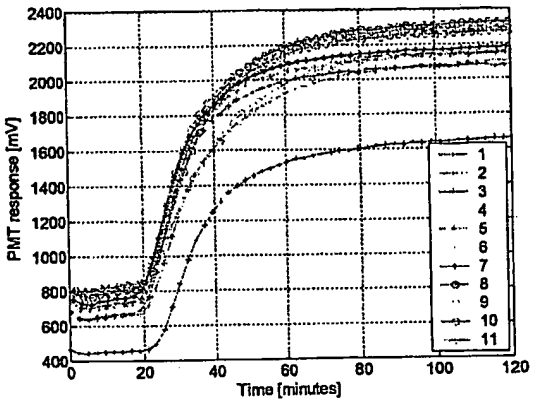

Figure 8

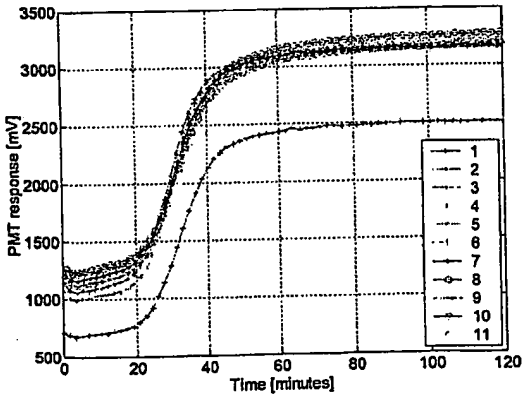

Figure 9

PATENT COOPERATION TREATY

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PCT

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(PCT Administrative Instructions, Section 411)

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Date of mailing *(day/month/year)* 14 March 2005 (14.03.2005)	
Applicant's or agent's file reference P60638WO00	**IMPORTANT NOTIFICATION**
International application No. PCT/GB05/000308	International filing date *(day/month/year)* 28 January 2005 (28.01.2005)
International publication date *(day/month/year)*	Priority date *(day/month/year)* 28 January 2004 (28.01.2004)
Applicant NORCHIP et al	

1. By means of this Form, which replaces any previously issued notification concerning submission or transmittal of priority documents, the applicant is hereby notified of the date of receipt by the International Bureau of the priority document(s) relating to all earlier application(s) whose priority is claimed. Unless otherwise indicated by the letters "NR", in the right-hand column or by an asterisk appearing next to a date of receipt, the priority document concerned was submitted or transmitted to the International Bureau in compliance with Rule 17.1(a) or (b).

2. *(If applicable)* The **letters "NR"** appearing in the right-hand column denote **a priority document which, on the date of mailing of this Form, had not yet been received by the International Bureau** under Rule 17.1(a) or (b). Where, under Rule 17.1(a), the priority document must be submitted by the applicant to the receiving Office or the International Bureau, but the applicant fails to submit the priority document within the applicable time limit under that Rule, **the attention of the applicant is directed** to Rule 17.1(c) which provides that no designated Office may disregard the priority claim concerned before giving the applicant an opportunity, upon entry into the national phase, to furnish the priority document within a time limit which is reasonable under the circumstances.

3. *(If applicable)* **An asterisk (*)** appearing next to a date of receipt, in the right-hand column, denotes **a priority document submitted or transmitted to the International Bureau but not in compliance with Rule 17.1(a) or (b)** (the priority document was received after the time limit prescribed in Rule 17.1(a) or the request to prepare and transmit the priority document was submitted to the receiving Office after the applicable time limit under Rule 17.1(b)). Even though the priority document was not furnished in compliance with Rule 17.1(a) or (b), the International Bureau will nevertheless transmit a copy of the document to the designated Offices, for their consideration. In case such a copy is not accepted by the designated Office as the priority document, Rule 17.1(c) provides that no designated Office may disregard the priority claim concerned before giving the applicant an opportunity, upon entry into the national phase, to furnish the priority document within a time limit which is reasonable under the circumstances.

Priority date	Priority application No.	Country or regional Office or PCT receiving Office	Date of receipt of priority document
28 January 2004 (28.01.2004)	0401868.5	GB	02 March 2005 (02.03.2005)

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